

Overview

Useful For

Providing additional information, which aids in the identification of hemoglobin variants

Method Name

Only orderable as part of a profile. For more information see:

- HAEV1 / Hemolytic Anemia Evaluation, Blood
- HBEL1 / Hemoglobin Electrophoresis Evaluation, Blood
- MEV1 / Methemoglobinemia Evaluation, Blood
- REVE2 / Erythrocytosis Evaluation, Blood
- THEV1 / Thalassemia and Hemoglobinopathy Evaluation, Blood and Serum

Cation Exchange High-Performance Liquid Chromatography (HPLC)

NY State Available

Yes

Specimen

Specimen Type

Whole Blood EDTA

Specimen Required

Only orderable as part of a profile. For more information see:

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- MEV1 / Methemoglobinemia Evaluation, Blood
- REVE2 / Erythrocytosis Evaluation, Blood
- THEV1 / Thalassemia and Hemoglobinopathy Evaluation, Blood and Serum

Specimen Minimum Volume

1 mL

Reject Due To

All specimens will be evaluated at Mayo Clinic Laboratories for test suitability.

Specimen Stability Information

Specimen Type	Temperature	Time	Special Container
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Whole Blood EDTA	Refrigerated	10 days	
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Clinical & Interpretive

Clinical Information

A large number of variants of hemoglobin have been recognized. Although many do not result in clinical or hematologic effects, clinical symptoms that can be associated with hemoglobin disorders include microcytosis, sickling disorders, hemolysis, erythrocytosis/polycythemia, cyanosis/hypoxia, anemia (chronic, compensated, or episodic), and increased methemoglobin or sulfhemoglobin results (M-hemoglobins).

For common, and many of the uncommon, hemoglobin variants, protein studies will be sufficient for definitive identification. High-performance liquid chromatography is a method that provides useful and supplementary information on most hemoglobin variants.

Reference Values

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Interpretation

This test is not interpreted in isolation, but as a part of a profile.

Cautions

Some hemoglobin disorders and variants, including common alpha-thalassemia conditions, are not detected by screening methods and require further reflex testing to identify. If there is a family history of a known hemoglobin disorder, prior therapy for a hemoglobin disorder, or otherwise unexplained lifelong/familial symptoms, such as hemolysis, microcytosis, erythrocytosis/polycythemia, cyanosis, or hypoxia are present, this should be clearly communicated to the laboratory so appropriate reflex testing can be added.

Recent transfusion may mask protein results, including hemoglobin electrophoresis, hereditary persistence of fetal hemoglobin by flow cytometry, stability studies, and sickle solubility studies depending on percentage of transfused cells present. Some hemoglobin variants can originate from the donor blood product and not from the tested recipient. These are typically found in low percentages.

If the patient has undergone a bone marrow transplant, the results may be atypical and should be interpreted in the context of clinical information.

Some therapies cause artefactual effects in protein studies, including hydroxyurea and decitabine (increased hemoglobin F levels), voxelotor (artefactual peaks), and gene therapy (alternate protein detection, beta T87Q, by mass spectrometry). Clear communication of prior therapy is strongly recommended.

Clinical Reference

1. Hoyer JD, Hoffman DR. The thalassemia and hemoglobinopathy syndromes. In: McClatchey KD, eds. Clinical Laboratory Medicine. 2nd ed. Lippincott, Williams and Wilkins; 2002:866-895
2. Szuberski J, Oliveira JL, Hoyer JD. A comprehensive analysis of hemoglobin variants by high-performance liquid chromatography (HPLC). *Int J Lab Haematol.* 2012;34(6):594-604
3. Van Delft P, Lenters E, Bakker-Verweij M, et al. Evaluating five dedicated automatic devices for haemoglobinopathy diagnostics in multi-ethnic populations. *Int J Lab Haematol.* 2009;31(5):484-495

Performance**Method Description**

Hemolysate of whole blood is injected into an analysis stream passing through a cation exchange column using high-performance liquid chromatography. A pre-programmed gradient controls the elution buffer mixture that also passes through the analytical cartridge. The ionic strength of the elution buffer is raised by increasing the percentage of a second buffer. As the ionic strength of the buffer increases, the more strongly retained hemoglobins elute from the cartridge. Absorbance changes are detected by a dual-wavelength filter photometer. Changes in absorbance are displayed as a chromatogram of absorbance versus time. (Huismann TH, Schroeder WA, Brodie AN, et al. Microchromatography of hemoglobins. III. A simplified procedure for the determination of hemoglobin A2. *J Lab Clin Med.* 1975;86:700-702; Ou CN, Buffone GJ, Reimer GL, Alpert AJ. High-performance liquid chromatography of human hemoglobins on a new cation exchanger. *J Chromatogr.* 1983;266:197-205; instruction manual: Bio-Rad Variant II Beta-thalassemia Short Program Instructions for Use, L70203705. Bio-Rad Laboratories, Inc; 11/2011)

PDF Report

No

Day(s) Performed

Monday through Friday

Report Available

Same day/1 day

Specimen Retention Time

Normal: 7 days; Abnormal: 14 days

Performing Laboratory Location

Rochester

Fees & Codes**Fees**

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- Authorized users can sign in to [Test Prices](#) for detailed fee information.
 - Clients without access to Test Prices can contact [Customer Service](#) 24 hours a day, seven days a week.
 - Prospective clients should contact their account representative. For assistance, contact [Customer Service](#).

Test Classification

This test has been modified from the manufacturer's instructions. Its performance characteristics were determined by Mayo Clinic in a manner consistent with CLIA requirements. This test has not been cleared or approved by the US Food and Drug Administration.

CPT Code Information

83021

LOINC® Information

Test ID	Test Order Name	Order LOINC® Value
HPLC	HPLC Hb Variant, B	No LOINC Needed

Result ID	Test Result Name	Result LOINC® Value
65615	HPLC Hb Variant, B	No LOINC Needed