

ESR1 Mutation Analysis, Next-Generation Sequencing, Tumor

#### Overview

#### **Useful For**

Assisting in the clinical management of patients with metastatic breast cancer by identifying tumors with evolving resistance to endocrine therapy

Stratifying prognosis of metastatic breast cancer

#### **Genetics Test Information**

This test uses targeted next-generation sequencing to evaluate for somatic mutations within the *ESR1* gene. See <u>Targeted Genes and Methodology Details for ESR1 Mutation Analysis</u> for details regarding the targeted gene regions evaluated by this test.

This test is performed to evaluate for somatic mutations within solid tumor samples. It **does not assess** for germline alterations within the *ESR1* gene.

#### **Additional Tests**

Test Id	Reporting Name	Available Separately	Always Performed
SLIRV	Slide Review in MG	No, (Bill Only)	Yes

# **Testing Algorithm**

When this test is ordered, slide review will always be performed at an additional charge.

## **Special Instructions**

- <u>Tissue Requirements for Solid Tumor Next-Generation Sequencing</u>
- Targeted Genes and Methodology Details for ESR1 Mutation Analysis

#### **Method Name**

Sequence Capture and Targeted Next-Generation Sequencing (NGS)

### **NY State Available**

Yes

## Specimen

## **Specimen Type**

Varies

# **Ordering Guidance**



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Multiple oncology (cancer) gene panels are available. For more information see <u>Hematology, Oncology, and Hereditary</u> Test Selection Guide.

# **Necessary Information**

A pathology report (final or preliminary), at minimum containing the following information, must accompany specimen for testing to be performed:

- 1. Patient name
- 2. Block number-must be on all blocks, slides, and paperwork (can be handwritten on the paperwork)
- 3. Tissue collection date
- 4. Source of the tissue

### Specimen Required

#### This assay requires at least 20% tumor nuclei.

NOTE: Submit tissue from either local recurrence or metastatic disease collected after endocrine therapy has been administered (see Clinical Information for more details).

- -Preferred amount of tumor area with sufficient percent tumor nuclei: tissue 216 mm(2)
- -Minimum amount of tumor area: tissue 36 mm(2)
- -These amounts are cumulative over up to 10 unstained slides and must have adequate percent tumor nuclei.
- -Tissue fixation: 10% neutral buffered formalin, not decalcified
- -For specimen preparation guidance, see <u>Tissue Requirement for Solid Tumor Next-Generation Sequencing</u>. In this document, the sizes are given as 4 mm x 4 mm x 10 slides as preferred: approximate/equivalent to 144 mm(2) and the minimum as 3 mm x 1 mm x 10 slides: approximate/equivalent to 36 mm(2).

# Preferred:

Specimen Type: Tissue block

Collection Instructions: Submit a formalin-fixed, paraffin-embedded tissue block with acceptable amount of tumor

tissue.

#### Acceptable:

**Specimen Type:** Tissue slides

Slides: 1 Stained and 10 unstained

**Collection Instructions**: Submit 1 slide stained with hematoxylin and eosin and 10 unstained, nonbaked slides with 5-micron thick sections of the tumor tissue.

**Note:** The total amount of required tumor nuclei can be obtained by scraping up to 10 slides from the same block.

Additional Information: Unused unstained slides will not be returned.

**Specimen Type:** Cytology slides (direct smears or ThinPrep)

Slides: 1 to 3 Slides

**Collection Instructions:** Submit 1 to 3 slides stained and coverslipped with a preferred total of 5000 nucleated cells, or a

minimum of at least 3000 nucleated cells.

Note: Glass coverslips are preferred; plastic coverslips are acceptable but will result in longer turnaround times.

**Additional Information**: Cytology slides will not be returned.

#### **Forms**

If not ordering electronically, complete, print, and send an Oncology Test Request (T729) with the specimen.



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# **Specimen Minimum Volume**

See Specimen Required

# Reject Due To

Specimens that	Reject
have been	
decalcified (all	
methods)	
Specimens that	
have not been	
formalin-fixed,	
paraffin-embe	
dded, except	
for cytology	
slides	
Extracted	
nucleic acid	
(DNA/RNA)	

# **Specimen Stability Information**

Specimen Type	Temperature	Time	Special Container
Varies	Ambient (preferred)		
	Refrigerated		

# **Clinical & Interpretive**

#### **Clinical Information**

The *ESR1* (estrogen receptor 1) gene encodes an estrogen receptor that regulates cell growth through activation of downstream signaling pathways upon binding of estrogen. Tumors demonstrating estrogen receptor expression by immunohistochemistry (ER-positive) are candidates for endocrine therapy, such as selective estrogen receptor modulators (SERM), selective estrogen receptor degraders/downregulators (SERD), and aromatase inhibitors. *ESR1* mutations are rarely observed in untreated breast cancers; however, mutations in the ligand-binding domain of *ESR1* can occur secondarily after exposure to aromatase inhibitors and other endocrine therapies in ER-positive metastatic breast tumors, frequently with multiple different mutations in *ESR1* occurring together. Current data suggests that *ESR1* mutations mediate resistance to endocrine therapy. Studies also suggest that *ESR1* mutations are an independent indicator of poor prognosis.

This test assesses for somatic mutations in *ESR1*, including the ligand-binding domain(exons 4-9 in reference transcript NM\_001122740). Breast cancers with mutations in the ligand binding domain of *ESR1* may be responsive to elacestrant (Orserdu), an endocrine therapy in the SERD class of drugs that is clinically approved for postmenopausal women or



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adult men with ER-positive, *HER2*-negative, *ESR1*-mutated advanced or metastatic breast cancer with disease progression following at least one line of endocrine therapy.

#### **Reference Values**

An interpretive report will be provided.

# Interpretation

The interpretation of molecular biomarker analysis includes an overview of the results and the associated diagnostic, prognostic, and therapeutic implications.

#### **Cautions**

This test cannot differentiate between somatic and germline alterations. Additional testing may be necessary to clarify the significance of results if there is a potential hereditary risk.

DNA variants of uncertain significance may be identified.

A negative result does not rule out the presence of a variant that may be present but below the limits of detection of this assay. The analytical sensitivity of this assay for sequence reportable alterations is 5% mutant allele frequency with a minimum coverage of 500X in a sample with 20% or more tumor content.

Point mutations and small deletion-insertion mutations will be detected in the *ESR1* gene only. This test may detect single exon deletions but does not detect multiexon deletions, duplications, or genomic copy number variants.

Variant allele frequency (VAF) is the percentage of sequencing reads supporting a specific variant divided by the total sequencing reads at that position. In somatic testing, VAF should be interpreted in the context of several factors including, but not limited to: tumor purity/heterogeneity/copy number status (ploidy, gains/losses, loss of heterozygosity) and sequencing artifact/misalignment.(1,2)

Rare polymorphisms may be present that could lead to false-negative or false-positive results.

The presence or absence of a variant may not be predictive of response to therapy in all patients.

Test results should be interpreted in the context of clinical, tumor sampling, histopathological, and other laboratory data. If results obtained do not match other clinical or laboratory findings, contact the laboratory for discussion.

Misinterpretation of results may occur if the information provided is inaccurate or incomplete.

Reliable results are dependent on adequate specimen collection and processing. This test has been validated on cytology slides and formalin-fixed, paraffin-embedded tissues; other types of fixatives are discouraged. Improper treatment of tissues, such as decalcification, may cause polymerase chain reaction failure.

### Supportive Data

Performance Characteristics:

The limit of detection for calling a somatic variant (single nucleotide variants [SNV] and deletions-insertions [delins, formerly indels]) is 5% variant allele frequency and having at least 500x deduplicated coverage.

Verification studies demonstrated concordance between this test and the reference method for detection of SNV and



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delins is 99.7% (699/701) and 96.6% (226/234) of variants, respectively. Concordance for the detection of delins was 98.9% (186/188) in variants 1 to10 base pairs (bp) in size, 95.8% (23/24) in variants 11 to50 bp in size, and 88.9% (8/9) in variants 51 to200 bp in size.

#### **Clinical Reference**

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- 2. Spurr L, Li M, Alomran N, et al. Systematic pan-cancer analysis of somatic allele frequency. Sci Rep. 2018;8(1):7735. Published 2018 May 16. doi:10.1038/s41598-018-25462-0
- 3. Arenedos M, Vicier C, Loi S, et al. Precision medicine for metastatic breast cancer-limitations and solutions. Nat Rev Clin Oncol. 2015;12(12):693-704. doi: 10.1038/nrclinonc.2015.123
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- 5. Gradishar WJ, Moran MS, Abraham J, et al. NCCN Guidelines Insights: Breast Cancer, version 4.2021. J Natl Compr Canc Netw. 2021;19(5):484-493. doi: 10.6004/jnccn.2021.0023
- 6. Toy W, Shen Y, Won H, et al. ESR1 ligand-binding domain mutations in hormone-resistant breast cancer. Nat Genet. 2013;45(12):1439-1445. doi: 10.1038/ng.2822
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- 9. Bidard FC, Kaklamani VG, Neven P et al. Elacestrant (oral selective estrogen receptor degrader) versus standard endocrine therapy for estrogen receptor-positive, human epidermal growth factor receptor 2-negative advanced breast cancer: results from the randomized phase III EMERALD trial. J Clin Oncol. 2022;40(28):3246-3256

## **Performance**

# **Method Description**

Next-generation sequencing is performed to evaluate the presence of a mutation in all coding regions of the *ESR1* gene. See <u>Targeted Genes and Methodology Details for ESR1 Mutation Analysis</u> for details regarding the targeted gene regions evaluated by this test.(Unpublished Mayo method)

A pathology review and macro dissection to enrich for tumor cells are performed prior to slide scraping

# **PDF Report**

No

#### Day(s) Performed

Monday through Friday

# **Report Available**

12 to 20 days



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# **Specimen Retention Time**

FFPE tissue block: Unused portions of blocks will be returned within 10-14 days after testing is complete; FFPE tissue/cytology slides: Unused tissue slides are stored indefinitely; Digital images are obtained and stored for all slides used in testing.

## **Performing Laboratory Location**

Rochester

#### **Fees & Codes**

#### **Fees**

- Authorized users can sign in to <u>Test Prices</u> for detailed fee information.
- Clients without access to Test Prices can contact <u>Customer Service</u> 24 hours a day, seven days a week.
- Prospective clients should contact their account representative. For assistance, contact <u>Customer Service</u>.

#### **Test Classification**

This test was developed and its performance characteristics determined by Mayo Clinic in a manner consistent with CLIA requirements. It has not been cleared or approved by the US Food and Drug Administration.

# **CPT Code Information**

88381 - Microdissection, manual 81479

# **LOINC®** Information

Test ID	Test Order Name	Order LOINC® Value
ESR1T	ESR1 Mutations Analysis, Tumor	102116-1

Result ID	Test Result Name	Result LOINC® Value
617929	Result	82939-0
617930	Interpretation	69047-9
617931	Additional Information	48767-8
617932	Specimen	31208-2
617933	Tissue ID	80398-1
617934	Method	85069-3
617935	Disclaimer	62364-5
617936	Released By	18771-6